Plasma Kinetics of a Chylomicron-Like Emulsion in Normolipidemic Obese Women After a Short-Period Weight Loss by Energy-Restricted Diet

M.R.M. Oliveira and R.C. Maranhão

Chylomicrons carry dietary fats in the bloodstream for storage in body tissues, and thus play an important role in obesity. The 2-step chylomicron metabolism consists of lipolysis by lipoprotein lipase (LPL) on vessel walls and hepatic uptake of triglyceride-depleted remnants. A triglyceride-rich emulsion that mimics chylomicrons, labeled with [9,10- 3 H]glycerol-trioleate (TG) and [1- 1 C] cholesteryl-oleate (CE) was intravenously injected into 14 obese women with body mass index between 30 and 40 kg/m² (age, 30 to 40 years), before and after a 2-month energy-restricted diet and into non-obese controls for determination of radioactive lipid plasma kinetics. TG kinetics evaluates lipolysis, whereas CE kinetics evaluates remnant removal. The emulsion TG fractional clearance rate (FCR, in min $^{-1}$) was similar in obese women and their controls (0.126 \pm 0.065; controls, 0.111 \pm 0.031), but the CE-FCR was pronouncedly reduced in the obese subjects (0.028 \pm 0.014; controls, 0.070 \pm 0.009 min $^{-1}$; P < .0001). After the energy-restricted diet, TG-FCR was reduced in the obese women (0.075 \pm 0.044 min $^{-1}$; P < .05), but CE-FCR was unchanged (0.032 \pm 0.025 min $^{-1}$). Therefore, the lipolysis of the chylomicron-like emulsion is normal in obese women, but remnant removal from the plasma is diminished. After active weight loss by an energy-restricted diet, the remnant removal was unchanged but lipolysis was diminished, possibly due to adaptative changes in LPL activity. *Copyright 2002, Elsevier Science (USA). All rights reserved.*

BESITY IS FREQUENTLY accompanied by alterations in plasma lipids, mainly by fasting hypertriglyceridemia.1,2 As measured after a 12-hour fasting period, concentration in the plasma of triacylglycerols mostly reflects the concentration of very-low-density lipoprotein (VLDL), the triacylglycerol-rich lipoprotein synthesized by the liver. VLDL accumulation in obese subjects may be due to saturation of the lipolytic process by excessive production of the lipoprotein. Increased VLDL synthesis by the liver is consequent to the enhanced supply of free fatty acids drained from the abdominal fat stores to the liver. This process is mediated by endocrine factors and high plasma concentration of insulin and cortisol are often found in subjects with the central form of obesity.³⁻⁵ Low plasma HDL-cholesterol may also be found in obesity consequent to high triacylglycerols. Because of the action of transfer proteins, net mass transfer of cholesterol from highdensity lipoprotein (HDL) to VLDL occurs when VLDL concentration increases. Low-density lipoprotein (LDL)-cholesterol may be increased, but in some studies this was not found.6,8

Chylomicrons are the triacylglycerol-rich lipoproteins synthe sized in the enterocyte from the fats and other lipids ingested in the diet and absorbed by the intestine. They share a common catabolic pathway with liver-produced VLDL. Similarly to VLDL, chylomicrons undergo the action of lipoprotein lipase (LPL) on the capillary wall, triggered by apolipoprotein CII (apo CII), one of the apolipoproteins present on the surface of the lipoprotein particles. Apo CII possesses a domain that binds to the enzyme and another that stimulates the enzymatic action. LPL can break chylomicron triacylglycerols down to glycerol and fatty acids, which are thus absorbed in several body tissues, especially muscle and adipose tissue, where the compounds are re-esterified and stored.^{9,10} This mechanism is important for the energetic economy of the organism, since fat constitutes its greatest storage energy source. By the action of hormonesensitive lipase, fats stored in the body tissues can again be broken down and released into the circulation for disposal by the liver when energy expenditure is required.^{9,10} After lipolysis by LPL, the resulting smaller particles, called chylomicron remnants, unbind from the enzyme molecules and return to circulation. They are then sequestered into the space of Disse and taken up by the liver cells by various receptor mechanisms, mainly the LDL receptors and LDL receptor-related protein (LRP). P-11 Apo E is the main ligand of chylomicron remnants to the hepatic receptors. Among the lipoprotein classes, chylomicrons possess the most rapid mechanism for plasma clearance, their half-life being about 15 to 20 minutes. As described above, the metabolism of chylomicrons is directly involved in the excess of fat stores that defines the obese organism.

Despite its crucial importance, only a few studies have addressed the issue of chylomicron metabolism in obese subjects. To shed more light on this issue, we studied a group of obese women before and after weight reduction was achieved by a 2-month energy-restricted diet period. In weight reduction programs, greater weight loss is usually achieved during this initial short period. Chylomicron metabolism was assessed using a triacylglycerol-rich emulsion model that mimics the intravascular behavior of chylomicrons. The emulsion is double-labeled with radioactive cholesteryl esters and triacylglycerols. After intravenous injection into the subjects, determination of the plasma decaying curves of the labeled lipids allows the 2-step metabolism of chylomicrons to be followed. The

From the Faculty of Pharmaceutical Sciences, and Heart Institute of the Medical School Hospital (INCOR-HCFMUSP), University of São Paulo, São Paulo, Brazil.

Submitted March 13, 2001; accepted March 12, 2002.

Supported by Grant No. 99/01229-2 from the the Fundação do Amparo à Pesquisa do Estado de São Paulo (FAPESP), São Paulo, Brazil. M.R.M.O received a scholarship from FAPESP and R.C.M. holds a Research Award from the Conselho Nacional de Desenvolvimento Científico e Tecnologico (CNPq), Brasília, Brazil.

Address reprint requests to R.C. Maranhão, MD, PhD, Laboratório de Lípides, Instituto do Coração do Hospital das Clínicas da FMUSP, Av. Enéas de Carvalho Aguiar, 44-CEP 05403-000 São Paulo-SP, Brazil

Copyright 2002, Elsevier Science (USA). All rights reserved. 0026-0495/02/5109-0004\$35.00/0 doi:10.1053/meta.2002.34698

1098 OLIVEIRA AND MARANHÃO

chylomicron-like emulsion model is a useful tool that facilitates human studies of this metabolic pathway. The triglyceride-rich emulsion approach has been validated in human subject studies. ^{14,15} It has revealed chylomicron metabolism defects in malignant hypertension, ¹⁶ systemic lupus erythematosus, ¹⁷ and heart transplant recipients, ¹⁸ and has been used to investigate effects of lipid-lowering drugs on chylomicron metabolism. ^{19,20} In this study, we also determined the plasma kinetics of the chylomicron-like emulsion in a control group of normal-weight women.

SUBJECTS AND METHODS

Subjects

Fourteen obese female volunteers with body mass index (BMI) between 30 and 40 kg/m² and aged 30 to 40 years participated in the study. Exclusion criteria were apparent or reported diseases, amenorrhea, pregnancy or breast feeding, alcohol abuse, use of antihyperlipidemia or anti-obesity medications, as well as dietary regimen for weight loss for the last 6 months. All participants were sedentary, were not smokers, and none had arterial hypertension. A control group consisting of 14 volunteer women aged 30 to 40 years with BMI less than 25 kg/m² was also selected according to the criteria described above.

The design and the objective of the study were explained to each participant before the study, and informed written consent was obtained from all. The study was approved by the Scientific and Ethics Committee of the Heart Institute of the Medical School Hospital of the University of São Paulo.

Weight Loss Treatment

The group of obese women was submitted to treatment for weight loss over a period of 2 months with the assistance of a multiprofessional team composed of a nutritionist, endocrinologist, and psychologist. Treatment was based on nutritional education and stimulation of changes in eating behavior. The participants were seen individually by the nutritionist once a week for dietary education and planning and were instructed to reduce energy and fat ingestion and to increase the ingestion of vegetables rich in fiber. The changes in eating habits were induced while respecting the sociocultural context and the limits of each individual. The control of food ingestion was performed using a daily eating record beginning the week that preceded the experiment.

Emulsion Preparation

The emulsion was prepared as previously described,²¹ with addition to the lipid mixtures of [1-¹⁴C]cholesteryl-oleate (CE; specific activity, 2.07 Gbq/mmol) and [9,10-³H]glycerol-trioleate (TG; specific activity, 518 GBq/mmol), supplied by Amersham International (Cardiff, UK). The emulsion was purified by 2-step ultracentrifugation, as described previously,²² sterilized by passage through a 0.2- μ m filter, and evaluated for sterility and pyrogenicity prior to injection into the patients.

Kinetics of the Emulsion

The determination of the plasma kinetics of the chylomicron-like emulsion was performed before the dietary intervention in both controls and obese subjects. In the 2 obese groups, the plasma kinetics test was repeated after the 2-month energy-restricted diet. The emulsion fraction injected in each subject had a lipid mass of approximately 3.0 mg in 500 μ L vol. [14 C] and [3 H] radioactivities of the labeled lipids were 74 kBq (2 μ Ci) and 148 kBq (4.0 μ Ci), respectively. The emulsion was injected by intravenous bolus after a 12-hour fast. Blood samples were collected from another peripheral vein at pre-established intervals over 45 minutes. Blood was centrifuged and the radioactivity

contained in 1.0 mL of plasma was measured by liquid scintillation counting (Packard 1.660 TR, Meridien, CT). The safety of the radioactive dose injected into the subjects was warranted according to radioprotection regulations²³ as described elsewhere.¹⁵

Compartmental Analysis

The removal of the emulsion from the plasma was evaluated by compartmental analysis according to a modification of the model proposed by Redgrave et al.14 Briefly, 4 compartments were employed to estimate the kinetic parameters for both ¹⁴C-CE and ³H-TG tracers. The plasma hydrolysis and removal of native chylomicrons, as well as chylomicron-like emulsions, displayed a rapid initial decay followed by a slow removal phase. 10,15,24 The $k_{x,y}$ constants represent the transfer or fractional catabolic rates (FCRs) from compartment x to compartment y. The kinetics of ¹⁴C-CE and ³H-TG metabolism are represented by compartments 1 to 4 and 5 to 8, respectively. The rapid and slow decay phases evaluated by the 14C-CE and 3H-TG tracers are represented by $k_{1,3}$ and $k_{2,3}$ and by $k_{5,7}$ and $k_{6,7}$, respectively. The model also takes into account the recirculation of the radioactive tracers in plasma in the form of newly synthesized VLDL (expressed by k3.4 and k7.8 for the ¹⁴C and ³H, respectively). The percentage of TG removed by the action of LPL was calculated from the differences between the areas under the curve for the removal of 14C-CE and of 3H-TG. All calculations were performed using a computer software.25 The details of the compartmental analysis calculations were published previously.20

Biochemical Analysis

Serum triacylglycerols, total, VLDL-, LDL-, and HDL-cholesterol, apo A1 and apo B, and glucose were determined from blood samples taken after a 12-hour fast using a automatic instrument (Cobas Mira Plus, Roche, Basel, Switzerland). Total cholesterol and triacylglycerols were determined with the aid of enzymatic test kits (CHO-PAD, Boehringer [Penzberg, Germany] and Abbott [Lake County, IL], respectively). HDL-cholesterol was determined with the same method, after precipitation of LDL and VLDL with MgCl₂ and phosphotungstic acid. VLDL- and LDL-cholesterol were calculated using the Friedewald equation.²⁶ The obese subjects were submitted to a glucose tolerance test before dietary intervention.

Statistical Analysis

All recorded variables were tabulated as means \pm SD or SEM. The differences in the obtained data were evaluated by the Student's t test, paired or unpaired when appropriate, with the level of significance set at P < .05 for all comparisons.

RESULTS

Plasma, Lipids, Apolipoproteins, and Glucose

Table 1 shows the individual physical characteristics and the plasma lipid and apoliprotein profiles of the obese women and their controls. Obese women had plasma lipid cholesterol and apo B and A1 values similar to the controls. Fast triacylglycerols showed a trend to be greater in the obese, but this was not statistically significant.

Fasting glucose plasma concentration in obese women was within the normal range (4.1 \pm 1.2 mmol). Postprandial glycemia determined 2 hours after the ingestion of the glucose load was below the 7.77-mmol/L cut-off value for glucose intolerance²⁷ in all but 1 obese woman.

Table 1. Individual Physical Characteristics and Plasma Lipid and Apoliprotein Profiles (mean ± SD) in Controls and Obese Women Before and 2 Months After an Energy-Restricted Diet

		Obese Women	
	Controls	Baseline	After Diet
Age (y)	37 ± 4	37 ± 5	
BMI (kg/m ²)	23.3 ± 0.98	$33.5\pm3.4*$	$30.9\pm4.2\$$
Weight (kg)	58.0 ± 4.70	85.9 ± 15.2*	79.7 ± 14.1§
Waist/hip ratio	0.75 ± 0.05	$0.84\pm0.06\dagger$	0.82 ± 0.07
Fasting glycemia			
(mmol/L)	3.88 ± 0.83	4.1 ± 1.2	4.1 ± 1.1
Postprandial glycemia			
(mmol/L)	4.33 ± 1.50	4.5 ± 1.6	
Triacylglycerols			
(mmol/L)	1.22 ± 0.62	1.49 ± 0.88	$1.17 \pm 0.76 \ddagger$
Cholesterol			
Total (mmol/L)	4.68 ± 0.62	4.62 ± 0.47	4.55 ± 0.46
VLDL (mmol/L)	0.56 ± 0.29	0.67 ± 0.41	$0.54 \pm 0.34 \ddagger$
LDL (mmol/L)	2.81 ± 0.60	2.82 ± 0.49	2.87 ± 0.52
HDL (mmol/L)	1.32 ± 0.33	1.14 ± 0.21	1.14 ± 0.21
Apo A1 (g/L)	1.72 ± 0.22	1.68 ± 0.43	1.59 ± 0.32
Apo B (g/L)	1.12 ± 0.18	0.95 ± 0.30	0.90 ± 0.29

^{*}P < .001, †P < .005 by unpaired Student's t test for controls in relation to the obese women at baseline.

Food Ingestion Before and During the Energy-Restricted Dietary Period

Table 2 lists data on the estimated dietary composition before and during the energy-restricted dietary period. During the week preceding the period of dietary intervention, there was a wide variation in the amount of ingested daily energy by the obese women, as calculated from their foodstuff intake records. Daily energy consumption was 8.36 ± 2.51 MJ/d (minimum, 4.48; maximum, 13.00 MJ/d). During the 2-month period of energy-restricted diet, the daily food intake was reduced to 4.60 ± 1.26 MJ/d, which corresponds to a 45% reduction in energy consumption.

Effect of the Energy-Restricted Diet on Anthropometric Measurements

After the 2-month dietary period, the obese women lost 7% of their initial weight. Figure 1 shows the week-to-week weight loss, expressed in grams. It is clear that there was a trend for diminished weight loss as the dietary period advanced. While

Table 2. Dietary Composition (mean ± SD) in Obese Women Before baseline and During the Energy-Restricted Diet Period

	Baseline	Energy-Restricted Diet
Energy (MJ/d)	8.36 ± 2.51	4.60 ± 1.26†
Fat (g/d)	73 ± 35	31 ± 12†
Protein (g/d)	93 ± 28	55 ± 19†
Carbohydrate (g/d)	240 ± 114	148 ± 46*

^{*}P < .05, †P < .005 by paired Student's t test for baseline in relation to the same group after diet.

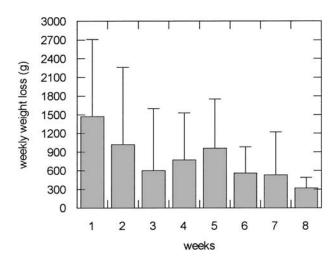


Fig 1. Weekly amount of weight loss during the 2-month dietary energy restriction period.

the weight loss was on average 1,500 g/wk at the first week of the dietary period, at the last week it was only 300 g/wk. The obese women showed reductions in all monitored anthropometric measurements (including triceps, biceps, subscapular, and suprailiac skinfolds; data not shown), except for the waist/hip ratio, which was unchanged (Table 1).

Effects of Energy-Restricted Diet on Plasma Lipids and Glucose

Table 1 shows the effects of the 2-month energy-restricted diet period on plasma lipids and apolipoproteins. Triacylglycerols and VLDL-cholesterol values were reduced. LDL- and HDL-cholesterol, as well as apo B, apo A1, and fasting plasma glucose, were unchanged.

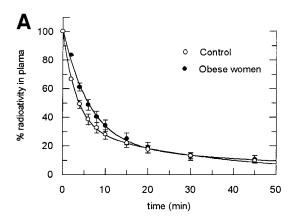
Emulsion Plasma Kinetics in the Obese

Figure 2 shows the plasma decay curves of the emulsion radioactive lipids obtained in obese women and their controls studied before the 2-month energy-restricted diet period. As expected, the decay curve of the emulsion triacylglycerol moiety was faster than that of the emulsion cholesteryl ester in all subjects. It is apparent that the triglyceride decay curves of the obese subjects and the controls were similar, but the cholesteryl ester curve of the obese subjects was slower than that of the controls.

Table 3 lists the plasma kinetic data calculated from the curves. TG-FCR was similar compared to the controls. CE-FCR, however, was smaller in the obese groups compared to the controls. Thus, the rate of triacylglycerols removed by lipolysis was estimated to be approximately 4-fold that of the controls. This difference in CE-FCR was largely due to the $k_{1\mbox{-}3}$ constant, which was 3-fold smaller in the obese subjects. According to the compartmental model used to analyze the data, the $k_{1\mbox{-}3}$ constant is related to the first exponential of the decay curve.

P < .005, P < .01 by paired Student's t test for baseline in relation to the obese women after diet.

1100 OLIVEIRA AND MARANHÃO



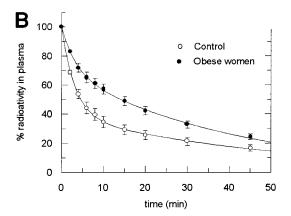


Fig 2. Removal from the plasma of the emulsion (A) [9,10-³H]glycerol-trioleate and (B) [1-¹⁴C]cholesteryl-oleate in the obese women and in the control normal-weight group. The emulsion labeled with the radioactive lipids was injected intravenously after a 12-hour fast. Plasma samples were taken at regular intervals over 45 minutes to determine the radioactivity remaining in the plasma in a scintillation solution.

Effects of Energy-Restricted Diet on Emulsion Kinetics

As shown Table 3 and Fig 3, TG-FCR was reduced by approximately 40% in obese women after the 2-month dietary period. Regarding triglyceride kinetics, the transfer constant $k_{6,7}$ was reduced, while $k_{7,8}$ was increased by the energy restriction, resulting in a net reduction of TG-FCR.

CE-FCR was apparently unchanged. Because TG-FCR was reduced by the diet, the rate of emulsion triacylglycerols removed by lipolysis was reduced. A 62% reduction was estimated.

DISCUSSION

Although not confirmed statistically, the obese women showed the trend for increase in fasting plasma triacylglycerols

Table 3. Plasma Kinetics Parameters of the Emulsion (mean ± SEM) in Obese Women Before and After 2 Months on an Energy-Restricted Diet

		Obese Women	
Parameters	Controls	Baseline	After diet
TG-FCR (min ⁻¹)	0.111 ± 0.031	0.126 ± 0.065	$0.075\pm0.044\$$
k _{5,6}	0.092 ± 0.031	0.138 ± 0.189	0.202 ± 0.023
k _{5,7}	0.305 ± 0.129	0.261 ± 0.308	0.231 ± 0.270
k _{6,7}	0.042 ± 0.018	0.061 ± 0.024	$0.032\pm0.013\S$
k _{7,8}	0.001 ± 0.001	0.001 ± 0.001	$0.003\pm0.003\S$
Lipolysis (%)	26.6 ± 9.65	70.0 ± 17.5*	$26.5 \pm 35.0 \ddagger$
CE-FCR (min ⁻¹)	0.070 ± 0.009	$0.028 \pm 0.014*$	0.032 ± 0.025
k _{1,2}	0.092 ± 0.031	0.138 ± 0.171	0.202 ± 0.122
k _{1,3}	0.277 ± 0.114	$0.076 \pm 0.130 \dagger$	0.157 ± 0.104
k _{2,3}	0.019 ± 0.002	0.021 ± 0.019	0.022 ± 0.024
k _{3,4}	0.001 ± 0.001	0.001 ± 0.001	0.000 ± 0.003

^{*}P < .0001, †P < .01 by unpaired Student's t test for controls in relation to the obese women at baseline.

Abbreviations: CE-FCR, [1-14C] cholesteryl-oleate fractional clearance rate; TG-FCR, [9,10-H₃] glycerol-trioleate fractional clearance rate.

that in other studies has been associated with obesity 1.6 and ascribed to VLDL overproduction by the liver, 28

The 7% weight loss obtained at the end of the 2-month dietary period confirms that the adherence of the participants to the program was good, similar to other studies that followed equivalent restriction dietary protocols. Weight reductions in the 5% to 10% range lead to changes in plasma lipids,^{29,30} mainly in fast triacylglycerols.^{31,32} In fact, after the dietary period, the obese women showed diminution of triacylglycerols. On the other hand, the weight loss had no effects on total, LDL-, or HDL-cholesterol, nor on the plasma concentrations of apo B and apo A1. This outcome is consistent with results of other studies in female subjects, where the alterations in LDL-and HDL-cholesterol after weight reduction were small or absent.^{33,34}

Among the methods to evaluate chylomicron metabolism, the use of double-labeled chylomicron-like emulsions injected into the bloodstream provides a useful study tool. Although devoid of apolipoproteins, the emulsions in contact with plasma acquire exchangeable apolipoproteins such as apo CII and apo E. Thus, the emulsion triacylglycerols undergo hydrolysis by LPL stimulated by apo CII and the resulting remnant particles are taken up by receptors in the liver that recognize apo E, similarly to chylomicron metabolic behavior. As documented previously,15 the cholesteryl ester component is not substantially removed from the emulsion to other lipoprotein density classes.35 Therefore, the radioactive cholesteryl ester is in fact the marker of the emulsion particles while in the intravascular compartment, and the finding of diminished CE-FCR indicates that the formed emulsion remnants were slowly removed from the circulation. This can be ascribed to the inhibitory effect of great fat ingestion on the receptor mechanisms that remove lipoproteins such as the LDL receptor, which also plays a major role in remnant removal.11

Because the TG-FCR was equal to that of the controls, it is presumed that the emulsion suffered greater lipolysis in the obese subjects, since lipolysis is estimated from the CE-FCR minus TG-FCR integration. Increased lipolysis rates in the

 $[\]ddagger P < .005, \$ P < .05$ by paired Student's t test for baseline in relation to the same group after diet.

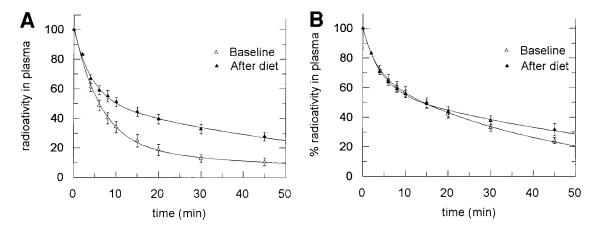


Fig 3. Removal from the plasma of obese women of the emulsion (A) [9,10-3H]glycerol-trioleate and (B) [1-14C]cholesteryl-oleate before and after the 2-month period of energy-restricted diet.

obese can be consequent to greater insulinemia associated with obesity because insulin stimulates the synthesis of LPL. 36

There are reports in the literature on postprandial lipidemia in obesity evaluated by the fat-load test. In this test, after the ingestion of a standard fatty meal to which vitamin A is added as a chylomicron particle marker, plasma triacylglycerols and vitamin A are measured over several hours. Lewis et al³⁷ observed that obese subjects had smaller rates of triacylglycerols removed than normal-weight subjects, but the removal of chylomicron remnants evaluated by the appearance-disappearance curve of vitamin A in the plasma was not altered. Vansant et al³⁸ also showed that obese women with an average BMI of 38 kg/m² had increased plasma triacylglycerols after a standard fatty diet, but their vitamin A curves were normal. Recently, Couillard et al39 and Taira et al40 reported both decreased post-prandial triacylglycerols and decreased vitamin A removal in subjects with high visceral adipose tissue compared to those with low visceral adiposity.

It is known that cholesteryl ester transfer protein (CETP) activity is enhanced in the obese. 41,42 By transferring cholesteryl esters from the emulsion to the plasma lipoprotein fractions that have intrinsically much smaller plasma clearances, enhanced CETP would conceivably decrease the FCR of the emulsion cholesteryl esters. However, in previous experiments we have shown that the CETP effect on the emulsion lipids is negligible, 15 probably due to the short plasma half-life of the emulsion, so that this hypothesis is unlikely to explain our results.

In patients with coronary artery disease or other conditions associated with coronary artery disease development, diminished removal of chylomicron-like emulsions and higher concentration of postprandial lipids in fat-load tests were reported. ^{15,16,43-46} This suggests that deficiencies in chylomicron intravascular catabolism are involved in atherogenesis. Remnant retention can therefore be related to increased atherosclerosis risk in obesity.

The effects on the chylomicron metabolism of the administration of an energy-restricted diet to obese women have not yet been documented. As our subjects had a less than 50-g/d weight reduction in the week preceding the post-diet

chylomicron-like emulsion test, they should be considered as being in a stable weight period.47 The results of the emulsion test showed diminution of lipolysis after the dietary period as estimated by the diminished emulsion triacylglycerol FCR and mainly by the diminished lipolysis index. In contrast, the fast triacylglycerol concentration did decrease after weight loss, indicating a lower VLDL concentration. This is conceivably due to diminished post-dietary period synthesis of this lipoprotein by the liver. 48,49 A number of factors could account for this metabolic response to the energy restriction. First, a decrease of LPL synthesis may follow dietary restriction. 50,51 As described by Taskinen and Nikkilä,52 during an energy-restricted diet period, LPL activity falls by 50% in adipose tissue and by 40% in skeletal muscle. This can be consequent to that diminished insulinemia that follows diminution of the caloric intake and the change of the chemical composition and physical form of carbohydrate in the dietary energy-restricted period,51,53,54 considering that LPL is insulin-dependent.36 The difference between the pre- and the post-diet period occurred in the second phase of the biexponential decay curve of the triglycerides, as seen in Fig 2A, involving k_{6.7} and k_{7.8}. An increase in k_{7.8} may also raise the possibility of post-diet increased recirculation of the emulsion lipids as newly synthesized VLDL.14,55

It is noteworthy that the removal of remnants, estimated by the emulsion cholesteryl ester, was not affected by the dietary restriction and weight loss.

In conclusion, obese women show increased emulsion lipolysis that diminishes after a short energy-restricted diet possibly due to the influence of diet-induced insulin level changes on LPL activity. Our results also showed a diminished ability to remove remnants, which did not improve after this short dietary period. Because delayed chylomicron remnant removal is associated with atherogenesis, ^{15,56-58} this finding can contribute to the increased incidence of coronary artery disease in obese subjects.

ACKNOWLEDGMENT

The authors are grateful to Dr Carlos H. Mesquita for the use of his Anacomp computational software for compartmental analysis.

1102 OLIVEIRA AND MARANHÃO

REFERENCES

- 1. Krotkiewski M, Björntorp P, Sjöström L, et al: Impact of obesity on metabolism in men and women: Importance of regional adipose tissue distribution. J Clin Invest 72:1150-1162, 1983
- 2. Lamon-Fava S, Wilson PWF, Schaefer EJ: Impact of body mass index on coronary heart disease risk factors in men and women: The Framingham Offspring Study. Arterioscler Thromb Vasc Biol 16:1509-1515, 1996
- 3. Jahr H, Ratzmann K-P, Beckert R, et al: Enhanced synthesis, storage, and secretion of insulin in pancreatic islets derived from obese subjects. Metabolism 32:1101-1106, 1983
- 4. Galanis DJ, McGarvey ST, Sobal J, eet al: Relations of body fat and fat distribution to the serum lipid, apolipoprotein and insulin concentrations of Samoan men and women. Int J Obes 19:731-738, 1995
- 5. Hautenen A, Adlercreutz H: Altered adrenocorticotropin and cortisol secretion in abdominal obesity: Implications for the insulin resistence syndrome. J Intern Med 234:461-469, 1993
- 6. Denke MA, Sempos CT, Grundy SM: Excess body weight: An under-recognized contributor to dyslipidemia in white american women. Arch Intern Med 154:401-410, 1994
- 7. Carlson LA, Lindstdt S: The Stockholm Prospective Study. Acta Med Scand 492:1-135, 1968 (suppl)
- 8. Seidell JC, Cigolini M, Charzewska J, et al: Fat distribution in european women: A comparation of anthropometric measurements in relation to cardiovascular risk factors. Int J Epidemiol 19:303-308, 1990
- Goldberg IJ: Lipoprotein lipase and lipolysis: Central roles in lipoprotein metabolism and atherogenesis. J Lipid Res 37:693-707, 1996
- 10. Hussain MM, Kancha RK, Zhou Z, et al: Chylomicron assembly and catabolism: Role of apoliproteins and receptors. Biochim Biophys Acta 1300:151-170, 1996
- Cooper AD: Hepatic uptake of chylomicron remnants. J Lipid Res 38:2173-2192, 1997
- 12. Linton MF, Hasty AH, Babaev VR, et al: Hepatic apo E expression is required for remnant lipoprotein clearance in the absence of the low density lipoprotein receptor. J Clin Invest 101:1726-1736, 1998
- 13. Redgrave TG, Maranhão RC: Metabolism of protein-free lipid emulsion models of chylomicrons in rats. Biochim Biophys Acta 835: 104-112, 1985
- 14. Redgrave TG, Ly HL, Quintão ECR, et al: Clearance from plasma of triacylglycerol and cholesteryl ester after intravenous injection of chylomicron-like emulsions in rats and man. Biochem J 290: 843-847, 1993
- 15. Maranhão RC, Feres MC, Martins MT, et al: Plasma kinetics of a chylomicron-like emulsion in patients with coronary artery disease. Atherosclerosis 126:15-25, 1996
- 16. Bernardes-Silva H, Toffoletto O, Bortolotto LA, et al; Malignant hypertension is accompanied by marked alterations in chylomicron metabolism. Hypertension 26:1207-1210, 1995
- 17. Borba EF, Bonfa E, Vinagre CG, et al: Chylomicron metabolism is markedly altered in systemic lupus erythematosus. Arthritis Rheum 43:1033-1040, 2000
- 18. Vinagre CG, Stolf NA, Bocchi E, et al: Chylomicron metabolism in patients submitted to cardiac transplantation. Transplantation 69:532-537, 2000
- 19. Santos RD, Spósito AC, Ventura LI, et al: Effect of prevastatin on plasma removal of chylomicron-like emulsion in men with coronary artery disease. Am J Cardiol 85:1163-1166, 2000
- 20. Santos RD, Ventura LI, Spósito AC, et al: The effects of gemfibrozil upon the metabolism of chylomicron-like emulsions in patients with endogenous hypertriglyceridemia. Cardiovasc Res 49: 456-465, 2001

- 21. Maranhão RC, Roland IA, Hirata MH: Effects of Triton WR 1339 and heparin on transfer of surface lipids from triglyceride-rich emulsions to high density lipoproteins in rats. Lipids 25:701-705, 1990
- 22. Redgrave TG, Roberts DCK, West CE: Separation of plasma lipoprotein by density-gradient ultracentrifugation. Anal Biochem 65: 42-49, 1975
- 23. Smith EM: Dose estimate techniques, in Rollo FD (ed): Nuclear Medicine Physics, Instrumentation and Agents. St Louis, MO, Mosby, 1977, pp 513-543
- 24. Redgrave TG, Zech LA: A kinetic model of chylomicron core lipid metabolism in rats: The effect of a single meal. J Lipid Res 5:473-482, 1987
- 25. Marchese SRM, Mesquita CH, Cunha IIL: Anacomp program application to calculate ¹³⁷C transfer rates in marine organisms and dose in man. J Radioanal Nucl Chem 232:233-236, 1998
- 26. Friedewald WT, Levy RI, Fredrickson DD: Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem 18:499-502, 1972
- 27. American Diabetes Association: Clinical practice recommendations: Screening for type 2 diabetes. Diabetes Care 23:S20-S23, 2000 (suppl)
- 28. Guerci B, Vergès B, Durlach V, et al: Relationship between altered postprandial lipemia and insulin resistance in normolipidemic and normoglucose tolerant obese patients. Int J Obes 24:468-478, 2000
- 29. Muls E, Kempen K, Vansant G, et al: The effects of weight loss and apolipoprotein E polymorphism on serum lipids, apolipoproteins A-I and B, and lipoprotein(a). Int J Obes Relat Metab Disord 17:711-716, 1993
- 30. Dattilo AM, Kris-Etherton PM: Effects of weight reduction on blood lipids and lipoproteins: A meta-analysis. Am J Clin Nutr 56:320-328, 1992
- 31. Andersen RE, Wadden TA, Bartlett SJ, et al: Regulation of weight loss to changes in serum lipids and lipoproteins in obese women. Am J Clin Nutr 62:350-357, 1995
- 32. Zimmerman J, Kaufmann NA, Fainaru M, et al: Effect of weight loss in moderate obesity on plasma lipoprotein and apolipoprotein levels and on high density lipoprotein composition. Arteriosclerosis 4:115-123, 1984
- 33. Thompson PD, Jeffery RW, Wing RR, et al: Unexpected decrease in plasma high lipoprotein cholesterol with weight loss. Am J Clin Nutr 32:2016-2021, 1979
- 34. Friedman CI, Falko JM, Patel ST, et al: Serum lipoprotein responses during active and stable weight reduction in reproductive obese females. J Clin Endocrinol Metab 55:258-262, 1982
- 35. Maranhão RC, Tercyak AM, Redgrave TG: Effects of cholesterol content on the metabolism of protein-free emulsion models of lipoproteins. Biochim Biophys Acta 875:247-255, 1986
- 36. Olivecrona T, Bergo M, Hultin M, et al: Nutritional regulation of lipoprotein lipase. Can J Cardiol 11:73G-78G, 1995 (suppl G)
- 37. Lewis GF, O'Meara NM, Soltys PA, et al: Postprandial lipoprotein metabolism in normal and obese subjects: Comparison after the vitamin A fat-loading test. J Clin Endocrinol Metab 71:1041-1050, 1000
- 38. Vansant G, Mertens A, Muls E: Determinants of postprandial lipemia in obese women. Int J Obes Relat Metab Disord 23:14-21, 1999 (suppl 1)
- 39. Couillard C, Bergeron N, Prud'homme D, et al: Postprandial triglyceride response in visceral obesity in men. Diabetes 47:953-960,
- 40. Taira K, Hikita M, Kobayashi J, et al: Delayed post-prandial lipid metabolism in subjects with intra-abdominal visceral fat accumulation. Eur J Clin Invest 29:301-308, 1999
 - 41. Arai T, Yamashita S, Hirano K, et al: Increased plasma cho-

lesteryl ester transfer protein in obese subjects. A possible mechanism for the reduction of serum HDL cholesterol levels in obesity. Arterioscler Thromb 14:1129-1136, 1994

- 42. Dullaart RP, Shiter WJ, Dikkeschei LD, et al: Effect of adiposity on plasma lipid transfer protein activities: A possible link between insulin resistence and ligh density lipoprotein metabolism. Eur J Clin Invest 24:188-194, 1994
- 43. Patsch JR, Miesenböck G, Hopferwieser T, et al: Relation of triglyceride metabolism and coronary disease: Studies in the postprandial state. Arterioscler Thromb 12:1336-1345, 1992
- 44. Cortener JA, Coates PM, Le N-A, et al: Kinetics of chylomicron remnant clearance in normal and in hyperlipoproteinemic subjects. J Lipid Res 28:195-206. 1987
- 45. Reznik Y, Pousse P, Herrou M, et al: Postprandial lipoprotein metabolism in normotriglyceridemic non-insulin-dependent diabetic patientes: Influence of apolipoprotein E polymorphism. Metabolism 45:63-71, 1996
- 46. Syvänne M, Hilden H, Taskinen M-R: Abnormal metabolism of postprandial lipoproteins in patients with non-insulin-dependent diabetes mellitus is not related to coronary artery disease. J Lipid Res 35:15-26, 1994
- 47. Schwartz RS, Brunzell JD: Increase of adipose tissue lipoprotein lipase activity with weight loss. J Clin Invest 67:1425-1430, 1981
- 48. Brunzell JD, Hazzard WR, Porte D, et al: Evidence for a comon, saturable, triglyceride removal mecanism for chylomicrons and very low density lipoprotein in man. J Clin Invest 52:1578-1585, 1973
 - 49. Karpe F, Steiner G, Olivecrona T, et al: Metabolism of triglyc-

- eride-rich lipoproteins during alimentary lipemia. J Clin Invest 91:748-758. 1993
- 50. Taskinen M-R, Nikkilä EA: Effects of caloric restriction on lipid metabolism in man. Atherosclerosis 32:289-299, 1979
- 51. Amri E-Z, Teboul L, Vannier C, et al: Fatty acids regulate the expression of lipoprotein lipase gene and activity in preadipose and adipose cells. Biochem J 314:541-546, 1996
- 52. Taskinen M-R, Nikkilä EA: Lipoprotein lipase of adipose tissue and skeletal muscle in human obesity: Response to glucose and to semistarvation. Metabolism 30:810-817, 1981
- 53. Pykalisto OJ, Smith PH, Brunzell JD: Determinants of human adipose tissue liprotein lipase: Effect of diabetes and obesity on basaland diet-induced activity. J Clin Invest 56:1108-1117, 1975
- 54. Slabber M, Barnard HC, Kuyl JM, et al: Effects of a low-insulinresponse, energy-restricted diet on weight loss and plasma insulin concentrations in hyperinsulinemic obese females. Am J Clin Nutr 60:48-53, 1994
- 55. Hultin M, Savonen R, Olivecrona T: Chylomicron metabolism in rats: Lipolysis, recirculation of triglyceride-derived fatty acids in plasma FFA, and fate of core lipids as analyzed by compartmental modelling. J Lipid Res 37:1022-1036, 1996
- 56. Barritt DW: Alimentary lipaemia in men with coronary artery disease and in controls. Br Med J 15:640-645, 1956
- 57. Zilversmit DB: Atherogenesis: A postprandial phenomenon. Circulation 60:473-485, 1979
- 58. Karpe F, Steiner G, Uffelman K, et al: Postprandial lipoproteins and progression of coronary atherosclerosis. Atherosclerosis 106:83-97, 1994